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# ANTIBACTERIAL, ANTIFUNGAL AND ANTIFEEDANT ACTIVITY OF QUINAZOLINONYL-β-LACTAMS/QUINAZOLINONES AND BIS (QUINAZOLINONYL-β-LACTAMS)

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#### **ABSTRACT**

Quinazolinonyl-β-lactams/quinazolinones, bis(quinazolinonyl-β-lactams) earlier synthesised by us are screened for antibacterial activity against *Staphylococcus aureus* and *Escherichia coli*, antifungal activity against *Fusarium oxisporium*, *Macrophomina sorgina* on PDA plates and antifeedant activity against agriculture pest *Achoea janata* were studied. All the compounds proved to be ineffective in antibacterial activity. The compounds 1, 2, 4, 9, 10 and 11 proved to be effective antifeedants. Compound 1 with chlorine substituent showed both antifungal & antifeedant activities.

**Keywords:** Quinazolinones,  $\beta$ -lactams, bis- $\beta$ -lactams, bisquinazolinones, antimicrobial, antifungal, antifeedant activity.

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#### **INTRODUCTION**

2-Heteryl and heteroalkyl-4(3H)-quinazolinones exhibited a wide range of pharmacological properties such as CNS depressant, antimicrobial, antibacterial, analgesic, antifungal, antiinflammatory, antiulcer, anticonvulsant, antihypertensive, sedative, anaesthetic, tranquilising and muscle relaxant, body temperature lowering, spore germination inhibition in *Drechslera rostrata* and *Fusarium oxysporum*, CNS active, hypnotic, antidepressant, antihelmentic, inhibition of AMPA receptor activation, antihistamine, virucidal, hypoglycemic, MAO inhibition, insecticidal, radioprotective, spasmolytic, contraceptive, antitubercular, antimonomine oxidase, H<sub>2</sub>-antagonist and antisecretion activity. They are also useful in the treatment of gastrointestinal and appetite disorders as cholecystokinin β-receptor and cholecystokinin gastric receptors. They also find application as heat stable epoxy resins, fiber reactive dyes and polymers. In view of these findings, we synthesised quinazolinonyl-β-lactams/quinazolinones, bis(quinazolinonyl-β-lactams) and tested for possible antibacterial, antifungal and antifeedant activities. The following compounds were synthesised in our laboratory and the detailed procedures were reported. 2-Chloromethyl-3-methyl-4 (3H)-quinazolinone (1)<sup>2</sup>, 2- (4-ethylphenyl)aminomethyl-3-methyl-4(3H)quinazolinone (2)<sup>2</sup>, 2-N-chloroacetyl 2 (4-methoxyphenyl)aminomethyl-3-methyl-4(3H)-quinazolinone  $(3)^2$ , 4-(3-methyl-4(3H)-quinazolinone-2-yl)-1-(4-methylphenyl)azetidin-2-one  $(4)^2$ , 4-(3-methyl-4(3H)quinazolinone-2-yl)-1-(4-ethoxyphenyl)azetidin-2-one (5)<sup>2</sup>, 4-(3-methyl-4(3H)-quinazolinone-2-yl)-1-(4-(**6** $)^2$ , 3,3'-dimethyl-4,4'-[*N*,*N*'-bis(4-(3-methyl-4(3*H*)-quinazolinone-2ethylphenyl)azetidin-2-one  $(7)^3$ , yl)methyl)|diaminobiphenyl 3,3'-dimethyl-4,4'-[N,N'-dichloroacetyl-N,N'-bis((3-methyl-4(3H)quinazolin-one-2-yl)methyl)|diaminobiphenyl  $(8)^3$ . 3.3'-dimethyl-4.4'-[bis(4-(3-methyl-4(3H)quinazolinone-2-yl)azetidin-2-one-1-yl)|biphenyl (9)<sup>3</sup>, 2-N-azidoacetylphenyl- aminomethyl-3-methyl-4(3H)-quinazolinone (10)<sup>4</sup>, 2-N-azidoacetyl(4-ethylphenyl) aminomethyl-3-methyl-4(3H)-quinazolinone  $(11)^4$ , 3'-acetamido-6'-chloro-3-methyl-2,2'-bis-4(3H)-quinazolinone  $(12)^5$ , 3'-acetamido-6'-bromo-3-

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methyl-2,2'-bis-4(3H)-quinazolinone (13)<sup>5</sup>, 3'-acetamido-6',8'-dichloro-3-methyl-2,2'-bis-4(3H)-quinazolinone (14)<sup>5</sup> were screened for their antibacterial, antifungal and antifeedant activities.

#### **EXPERIMENTAL**

# **Evaluation of antifungal activity Organisms and growth conditions**

Fusarium oxisporium and Macrophomina sorgina fungi were grown in medium (pH 5.0), comprising of 1.5% potato dextrose agar (PDA), 0.09% KH<sub>2</sub>PO<sub>4</sub>, 0.02% MgSO<sub>4</sub>.7H<sub>2</sub>O, 0.02% KCl, 0.1% NH<sub>4</sub>NO<sub>3</sub>, 0.0002% FeSO<sub>4</sub>, 0.0002% ZnSO<sub>4</sub> and 2% agar. All the constituents were mixed in double distilled water and autoclaved at 121°C (at 15lb pressure) for 15 minutes. The medium when cooled to about 50 to 45°C temperature mixed with fungal strains separately and poured into sterile disposable petri plates for solidification of the medium. The PDA plates thus prepared were then kept for antifungal testing.

#### Antifungal testing<sup>6</sup>

The above mentioned PDA plates inoculated with two different fungal strains were taken and labeled separately. Sterile filter paper discs (6 mm diameter) prepared from standard Whatman No.1 filter paper was applied with test solution of different concentrations. After drying the discs were introduced on to the above inoculated PDA plates containing fungal strains. The plates with test compound discs were incubated for 3 days at 30°C.

After 3 days of the treatment, petri dishes were checked for growth inhibition zone. The presence or absence of growth inhibition zone around each disc was recorded by comparing with control (acetone). Presence of clear zone around the disc indicated the inhibition of fungal growth. The compound was then considered to be active against the fungi and the area of inhibition zone was measured. If no clear zone was observed around the disc, it indicated the inactivity of the test sample. The inhibition zone was calculated and compared with the control (untreated check).

#### **Evaluation of Insect-antifeedant activity:**

#### **Organism**

The test insect *Achoea janata* was maintained under laboratory conditions of  $27 \pm 1$   $^{0}$ C and  $70 \pm 5\%$  RH. The test larvae (IV instar) were fed on natural food - castor leaves. The antifeedant activity was assessed by using leaf discs in a "non-choice test method of Ascher and Rones.<sup>7</sup>

#### **Antifeedant Test**

Circular leaf discs of 9 cm diameter were cut from fresh castor leaves and treated with the solution of test compound from 100 to  $3.125\mu g/cm2$  by using serial dilutions. These leaf discs were air dried for 2-5 sec in acetone and were kept in petri dishes. Control discs were treated with acetone and Carbindazime and kept in separate petri dishes. Preserved IV instar larvae of *Achoea janata* were released simultaneously into petri dishes.

The consumption of leaf by the insect was measured after 48h. The leaf area consumed by the insect (in both control and treated) was measured by planimeter and the percentage of protection (antifeedant activity) was assessed by Singh and Panth formula.<sup>8</sup>

% of antifeedant = 
$$\frac{\left(\% \text{ of protection in treated}\right) - \left(\% \text{ of protection in control}\right)}{100 - \left(\% \text{ of protection in control}\right)} \times 100$$

### **Evaluation of antibacterial activity**

#### **Organisms**

The bacterial strains used in the experiments were *E. coli* of MTCC 119 (Gram -ve) and *S. aureus* of MTCC 96 (Gram +ve) and were procured from IMTECH, Chandigarh, India.

#### **Preparation of sample solution**

The test compounds were dissolved in acetone (AR grade) and prepared into different concentrations by successive dilutions. Whatman No.1 filter paper discs were dipped in different sample solution and kept for air drying for further use in testing for bacteriological activity. The concentrations of test compounds ranged from 50 to 1.625µg/disc.

#### Preparation of medium

The medium was prepared by adding agar powder (Thomas Baker) to nutrient broth, which was prepared with the beef extract known Lablemco (10 g), sodium chloride (5 g) and peptone (10 g) in water (1 L) and pH was adjusted to 7.2 - 7.4 and sterilised in autoclave at  $121^{\circ}$ C (at 15 lb pressure) for 15 minutes. The sterile nutrient agar medium was cooled down to  $45-50^{\circ}$ C and poured into sterile petri plates of 6" diameter with a volume of 20 mL per plate and kept for solidification. The sterile agar plates were used for inoculating the bacterial cultures.

#### **Bacteriological testing**

Actively growing agar slant culture suspension of bacteria swab was inoculated separately on these solidified agar plates. Sterile filter papers were dipped in the test solution of different concentrations in acetone solvent (0.01 mL/disc). After drying the discs, they were introduced on to the above inoculated agar plates containing bacterial strains. The plates with test compound discs were incubated for 24h at  $37^{\circ}$ C. After 24h, the petri dishes were checked for growth inhibition zone. The presence or absence of growth inhibition zone around each disc was recorded by comparing with inhibition zone of standard disc (Streptomycin,  $10 \,\mu\text{g/disc}$ ). Presence of clear zone around the disc indicated the inhibition of organism's growth and measured the area of inhibition zone. The compound was then considered to be active against the organism. If no clear zone was observed around the disc, it indicated the inactivity of the test sample.

#### RESULTS AND DISCUSSION

#### **Antifungal activity**

Fusarium oxisporium and Macrophomina sorgina fungi were used to evaluate the antifungal activity of the test compounds. The experimental results and assessment of antifungal activity of the compounds are given in the Table-1.

#### **Antifeedant activity**

After 48h, antifeedant activity of compounds against *Achoea janata* was measured by using the planimeter and protected area (antifeedant activity) was calculated by using Singh and Panth formula. Likewise, all the experiments were carried out in triplicate and the average is reported in table – II.

#### **Antibacterial activity**

The above mentioned compounds were tested for antibacterial activity by using inhibition zone method on agar plates<sup>9</sup> against *Escherichia coli* (Gram -ve) and *Staphylococcus aureus* (Gram +ve). None of the compounds showed antibacterial activity.

#### **CONCLUSION**

#### **Antifungal activity**

Nine out of fourteen compounds showed most promising antifungal activity against Fusarium oxisporium at  $100\mu g/disc$  concentration and four compounds were fairly active against Macrophomina sorgina at  $12.5\mu g/disc$  concentration. In general compound 1 has exhibited ~83.00% antifungal activity against both fungal strains at  $50 \mu g/disc$  concentration. Compound 12 has exhibited 70.25% antifungal activity against Macrophomina sorgina at  $25\mu g/disc$ .

#### **Antifeedant activity**

All the compounds tested (1 to 14) exhibited moderate to good antifeedant activity against *Achoea janata* at 200 to  $6.25\mu g/cm^2$  concentrations. The activity persisted even after 48h, though it generally recedes after 24h. Of the fourteen compounds tested, 3,3'-dimethyl-4,4'-[N,N'-dichloroacetyl-N,N'-bis((3-methyl-4(3H)-quinazo- linone-2-yl)methyl)]diaminobiphenyl (8) is the most promising with 66.26% antifeedant activity even at  $6.25\mu g/cm^2$  concentration, and can be considered as a potent antifeedant. SAR studies of

**8** are preserved for future studies. Compounds **1**, **2**, **4**, **8**, **9** and **10** showed more than 50% leaf protection at  $6.25\mu g/cm^2$ .

#### **Antibacterial activity**

None of the compounds showed antibacterial activity.

Fig.-1: Structures of Different compounds

Table-1: Antifungal activity of compounds

	Fusarium oxisporium						Macrophomina sorgina					
Com	Concentration*					Concentration*						
	100 3.125	50	25	12.5	6.25		100	50	25	12.5	6.25	3.125
1			39.65	12.88			89.66	72.23	69.38	35.23	28.66	10.78
2	54.76	53.86					40.77	25.45	20.67	10.26		
3							40.77	35.45	30.67	10.26		
4	21.26											
5	21.26											
6	28.33	23.86										
7												
8												
9	32.86						39.66	22.86				
10	26.54	9.34										
11	13.54											
12	28.96	19.88					82.3	70.28		52.86	22.39	
13							61.86	55.58	48.33	25.62	7.23	
14	16.77						61.87	49.26	27.60	8.25		
TC	96.25	92.33	78.86				98.87	96.33	89.89	76.38	64.88	58.66
UTC												

Table-2: Antifeedant activity of compounds against Achoea janata after 48 hrs of treatment

Comp	% of Antifeedant activity after 48 hrs of treatment								
No.	Concentration in µg/cm <sup>2</sup>								
	200	100	50	25	12.5	6.25			
1.	82.70±8.62	81.36±5.00	60.39±2.90	58.13±2.86	53.72±4.31	55.96±2.75			
	(65.82)	(64.55)	(51.00)	(49.68)	(47.14)	(48.43)			
2.	71.76±0.64	67.88±3.06	60.46±3.06	53.94±3.86	52.67±4.05	50.73±3.34			
	(57.90)	(55.49)	(51.05)	(47.27)	(46.54)	(45.42)			
3.	68.11±1.29	45.65±3.08	26.20±4.48	21.40±2.53	15.21±0.55	11.28±1.70			
	(55.62)	(42.50)	(30.73)	(27.53)	(22.96)	(19.59)			
4.	68.41±1.01	65.32±4.60	63.75±1.77	57.43±3.22	53.66±3.84	51.29±3.21			
	(55.80)	(53.94)	(52.98)	(49.28)	(47.11)	(45.74)			
5.	72.98±7.05	70.70±2.20	60.94±3.67	49.26±1.40	36.97±4.46	37.76±2.39			
	(58.80)	(57.28)	(51.33)	(44.55)	(37.43)	(37.91)			
6.	62.90±2.51	58.74±4.71	60.50±2.78	54.44±3.02	45.04±4.75	43.25±6.08			
	(52.48)	(50.05)	(51.06)	(47.55)	(42.15)	(41.10)			
7.	61.96±5.45	60.73±2.65	60.33±1.16	48.01±3.77	44.11±1.71	38.33±2.64			
	(51.95)	(51.20)	(50.96)	(57.72)	(41.62)	(38.24)			
8.	88.99±2.40	84.64±2.46	82.51±4.14	78.37±1.80	70.67±3.26	66.26±0.83			
	(66.45)	(63.93)	(65.37)	(62.29)	(57.23)	(54.490)			

Concentration expressed in µg/cm² dissolved in Acetone No inhibition of the fungal growth (i.e., compound is not antifungal)

<sup>(-)</sup> UTC TC Untreated check (acetone)

Treated check (carbindazime)

9.	73.34±1.87	68.42±4.71	66.71±3.20	62.92±3.24	57.30±1.03	53.12±0.64
	(58.92)	(55.84)	(54.77)	(52.50)	(49.20)	(46.79)
10.	91.70±1.88	91.52±1.56	65.39±0.98	61.01±3.22	59.55±2.54	53.18±4.14
	(73.32)	(73.11)	(53.96)	(51.36)	(50.51)	(46.83)
11.	90.73±2.54	74.25±3.50	17.44±2.44	10.00±1.77	8.02±2.01	5.60±0.84
	(72.39)	(59.54)	(24.65)	(18.79)	(16.37)	(13.66)
12.	89.88±4.52	63.62±4.94	63.27±2.56	54.17±5.39	46.67±3.12	39.80±1.80
	(71.77)	(53.93)	(52.70)	(47.41)	(43.09)	(39.11)
13.	86.18±2.92	81.05±1.93	55.21±4.50	43.25±2.39	38.79±1.50	29.23±1.84
	(68.25)	(64.21)	(48.00)	(41.12)	(38.52)	(32.72)
14.	59.44±2.69	60.92±2.16	47.27±0.74	44.24±3.07	44.82±4.46	42.36±1.77
	(50.45)	(51.31)	(43.43)	(41.69)	(42.02)	(40.60)
UTC.	0.00	0.00	0.00	0.00	0.00	0.00
	(4.05±0.00)	(4.05±0.00)	(4.05±0.00)	(4.05±0.00)	(4.05±0.00)	(4.05±0.00)

UTC: untreated control

Significance at p<0.01.

SEM 9.47

CD at 0.05%; CV(%) 5.56

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#### REFERENCES

- 1. P.S.N. Reddy, P.P. Reddy and T. Vasantha, Heterocycles, 60, 183 (2003).
- 2. P.S.N. Reddy, Ch. Nagaraju and T. Vasantha., *Indian. J. Chem*, **38B**, 40 (1999).
- 3. P.S.N. Reddy, P.P. Reddy and T. Vasantha, *Indian. J. Chem*, **41B**, 1946 (2002).
- 4. P.S.N. Reddy, P.P. Reddy and T. Vasantha, *Indian. J. Chem*, **42B**, 393 (2003).
- 5. P.S.N. Reddy, P.P. Reddy and T. Vasantha, *Indian. J. Chem*, **41B**, 1950 (2002).
- 6. A. Sivan and I.Chit, J. General Microbiology, 135, 675 (1989).
- 7. K.S. R. Ascher and G. Rones, *International. Pest. Control*, 6 (1964).
- 8. R.P. Singh and N.C. Panth, *Experientia*, **36.** 552 (1980).
- 9. R. Cruickshank, J.P. Duguid, B.P. Marimion and R.H.A. Swain, *Medical Microbiology-2*, Churchil Livingstone, Edenburge, 190 (1975).

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<sup>\*</sup> Values in the parenthesis are transformed values (Arc sine)